



Phytoremediation of Heavy Metal-Contaminated Soils: An Overview of Principles and Expectations for Fundamental Techniques

Ashenafi Nigussie^{1, *}, Haymanot Awgchew²

¹Department of Natural Resource Management, Wondo Genet Agriculture Research Center, Shashemene, Ethiopia

²School of Plant and Horticultural Sciences, Debre Berhan University, Debre Berhan, Ethiopia

Email address:

nashenafinew@gmail.com (A. Nigussie)

*Corresponding author

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Abstract: The earth is currently dealing with a variety of issues and is losing its potential as a result of climate change brought on by increasing industrialization and urbanization. Harmful metals wastes generated by anthropogenic processes such as household, municipal, agricultural, industrial, and military operations penetrate the soil, decreasing its quality and usefulness. Because soil is the foundation of life, it necessitates excellent remediation activity. The problem of soil pollution is no longer being ignored because it is limited or no new land to replace. Therefore, the objective of this review paper is to explore the concepts and promises of basic phytoremediation approaches for heavy metal-contaminated soils. The use of living organisms, particularly plants (phytoremediation), is one of the remediation approaches that is now being used. In comparison to other soil remediation approaches, phytoremediation is an effective and affordable technology that can work with few maintenance costs once established, is suited for vast regions with low to moderate amounts of contaminants, and is ecologically benign. Phytoremediation, on the other hand, is a long-term remediation option, and not all of its remediation procedures are optimal. For example, in the case of phytovolatilization, air pollution may occur, while in the case of phytoextraction, pollutants collected in leaves may be released back into the environment during litterfall. Therefore, future concerns should be directed toward the modification and improvement of phytoremediation technologies that are likely to improve metal-binding abilities in plant tissues and phyto-transform toxic metals. Finally, it is critical to minimize or avoid the release of harmful compounds into the environment, in addition to enhancing and adapting various techniques.

Keywords: Heavy Metal, Pollutant, Phytoremediation, Toxicity

1. Introduction

Heavy metals are conventionally known as natural elements with metallic properties (ductility, conductivity, stability as cations, ligand specificity, etc), higher atomic weights, and a density five times greater than that of water [60, 115]. The most common and important heavy metals are As, Sr, Cs, U, Cd, Cr, Cu, Hg, Pb, and Zn [63, 96]. Some of these metals such as Zn, Cu, Mn, Ni, and Co are micronutrients necessary for plant growth and development while others (Cd, Pb, and Hg) have unknown biological functions [42]. As stated by Jayanthi *et al.* [56]; Shazia *et al.* [106], heavy metals are not

only cytotoxic but also carcinogenic and mutagenic in nature even when present at trace amount [56, 106]. For instance, the U.S Environmental Protection Agency and International Agency for Research on Cancer declared certain heavy metals like Pb, Cd, Hg, As and Cr as the most toxic of all pollutants and termed them carcinogens due to their potential problems to human health [115].

Electronics, automotive, ceramics, glass, paints, pigments, fertilizers, reagents, alloys for diverse uses, and so on are examples of heavy metal resources attributed to the fundamental acquired usefulness that are produced directly or indirectly from heavy metals. As a result, human

socioeconomic progression is becoming increasingly reliant on the use of various heavy metals, necessitating the extraction of enormous quantities of these resources as industrialization and urbanization progress [73]. However, most human activities emit non-recovered heavy metal-containing pollutants into the soil on a daily basis [78]. As a result, their negative consequences on human health and the environment are rapidly expanding [31] athering of minerals,

erosions, and volcanic activity are the most significant natural sources, while mining, smelting, electroplating, excessive use of pesticides, fertilizer discharge, biosolids (livestock manures, composts, and municipal sewage sludge), atmospheric deposition, and other anthropogenic sources are also significant [38, 99]. To be sure, heavy metal pollution and its effects on environmental quality and human health are well-known global issues [20, 105].

Table 1. Potential sources of heavy metals.

Heavy Metal	Sources	References
Arsenic*	Mining, smelting, pesticides, bio-solids, wood preservatives, petroleum refining, food additives, coal-based power plants, volcanic eruptions.	[31, 37]
Mercury*	Volcanic activities, waste from caustic soda industry, Gold-Silver mining, medical waste, peat, burning of wood and coal.	[31, 36]
Lead*	Petroleum derivatives, mining, paints, smelting, industrial and municipal sewage, pesticides, wastes from batteries,	[31, 73]
Cadmium*	Smelting, sludge, combustion of fossil fuels, phosphate fertilizers, paint, pigments, plastic stabilizers, electroplating.	[31, 36]
Chromium*	Electroplating, sludge, solid wastes, fly ash, tanning, textile, steel and pulp processing industries.	[31, 73]
Copper**	Copper polishing, mining, paint, plating, printing operations	[31, 36]
Nickel**	Electroplating, non-ferrous metal, paints, porcelain enameling	[36, 23]
Selenium**	Coal combustion, mining	[31]
Silver**	Battery manufacture, mining, photographic processing, smelting	[85, 84]

Source: Ayansina and Olubukola, [10] **; Mukhtar *et al.* [79] *

Several strategies for cleaning up the environment from these types of toxins are already in use, but the majority of them are expensive and fall short of their potential. Chemical procedures produce significant volumes of sludge, which raises expenses [87]; chemical and thermal processes are technically challenging and costly, and they can destroy soils' valuable components [50]. Traditionally, heavy metal contaminated soils have been remedied by either onsite management or excavation and subsequent disposal to a landfill. This type of disposal only relocates the contamination problem, as well as the risks associated with transporting polluted soil and contaminant migration from the landfill into the surrounding ecosystem. Soil washing is an alternative to excavation and landfill disposal for clearing polluted soil. However, this process is expensive and produces a heavy metal-rich residue that will require further treatment. Furthermore, because these physio-chemical procedures for soil remediation remove all biological activity, they render land unusable as a substrate for plant growth [42].

Concerns about environmental pollution have prompted the development of devices to determine the presence and mobility of metals in soil [108]. Phytoremediation is now an effective and cost-effective technological technique for removing metal contaminants from polluted soil. Because the expenses of cultivating a crop are low compared to the costs of soil removal and replacement, plant-based remediation solutions can operate with little upkeep once established. Phytoremediation is ten times cheaper than engineering-based remediation technologies such soil excavation, soil washing or burning, or pump-and-treat systems because biological processes are ultimately solar-driven [41].

The fact that phytoremediation is done in situ adds to the cost-effectiveness of the process and may limit the amount of polluted substrate that is exposed to humans, wildlife, and the

environment [82]. It is not, however, always the greatest answer to a contamination issue. The application of phytoremediation is limited by the meteorological and geological characteristics of the cleaning site, such as temperature, altitude, soil type, and agricultural equipment accessibility [100]. Furthermore, soil factors such as pH, organic matter, and clay concentration influence heavy metal bioavailability [8]. The use of a phytoremediation strategy may cause some issues. Woods that accumulate contaminants, for example, might be utilized as fuel, while pollutants gathered in leaves can be released back into the ecosystem during litterfall [100]. Many of the limits of phytoremediation can be summarized as follows: pollutants must be accessible to a plant and its root systems [82].

The problem of soil pollution is no longer being ignored because it is limited or no new land to replace. Furthermore, it is self-evident that the "used up" soil resource must be refined in an environmentally responsible and cost-effective manner in order to sustain life on Earth. Therefore, the major objective of this review paper is to explore the concepts and promises of basic phytoremediation approaches for heavy metal-contaminated soils.

2. Phytoremediation of Soils Polluted with Heavy Metals

2.1. An Overview of Phytoremediation Technology

Phytoremediation is a newly evolving field of science and technology that uses plants and their associated rhizospheric microorganisms to extract, sequester, and/or detoxify a wide variety of environmental contaminants [72]. A set of ecological techniques that employ plants in situ to promote pollutant breakdown, immobilization, and removal from the environment. It is a green technology that is

frequently the most cost-effective treatment for metal-polluted soils, especially in cases of widespread pollution [27]. Because many cropping cycles may take several years to decrease metals to acceptable regulatory levels, phytoremediation should be seen as a long-term remediation strategy.

Plants have a unique ability to concentrate elements and compounds from the soil and metabolize the molecules in their tissues, which is used in phytoremediation [45]. Plant roots can play a key role in metal removal via filtration, adsorption, and cation exchange, as well as plant-induced chemical changes in the rhizosphere, because the majority of plant roots are located in the soil [32, 129]. Physiological adaptations also regulate hazardous metal accumulations in

the roots by sequestering metals [43]. Variations in plant species, plant growth stage, and element characteristics may all influence metallic pollutant absorption, accumulation, and translocation.

The greatest progress in phytoremediation has been made with heavy metals [17, 128]. Soil-focused phytoremediation technologies are suitable for large areas which have been contaminated with low to moderate levels of contaminants. Phytoremediation will not be able to remediate extremely contaminated sites since the harsh conditions will prevent plant development and survival. The amount of soil that can be cleansed or stabilized is limited to the root zone of the plants. This depth might range from a few inches to many meters, depending on the plant [102].

Table 2. List of plant species utilized for phytoremediation of heavy metals.

Heavy Metal	Plant Species	References
Fe, Al, Cu, Mn, Cr, As, Zn, Hg	Jatropha (<i>Jatropha curcas</i> L.) **	[55, 131]
Cu, Fe, Mn, Zn, Ni, Cd, Pb, Co, As	Lettuce (<i>Lactuca sativa</i> L.) **	[3, 86, 88]
Pb, Cu, Zn, Fe, Cd, Ni, As, Cr	Pea (<i>Pisum sativum</i> L.) **	[39, 49, 66, 103, 127]
Cd, Cu, Fe, Ni, Pb, Zn, Cr	Spinach (<i>Spinacia oleracea</i> L.) **	[1, 54, 80, 81, 95]
As, Cd, Fe, Pb, Hg	Cress (<i>Lepidium sativum</i> L.) **	[44, 111]
As, Cd, Fe, Pb, Cu	Radish (<i>Raphanus sativus</i> L.) **	[44, 47]
Cd, Cu, Pb, Zn	<i>Salix</i> spp. (<i>S. viminalis</i> , <i>S. fragilis</i>) *	[94, 123]
	<i>Populus</i> spp. (<i>P. deltoides</i> , <i>P. nigra</i> , <i>P. trichocarpa</i>)*	[94]
	<i>Brassica juncea</i> L.**	[11, 103, 109, 118]
	Canola (<i>Brassica napus</i>) **	[29, 107, 118]
	<i>Zea mays</i> L.**	[1, 117]
Cd, Cu, Ni, Pb	Jatropha (<i>Jatropha curcas</i> L.)*	[2, 55]
Cd, Pb, Cr, Cu	Chickpea (<i>Cicer arietinum</i> L.)**	[28, 57, 126]
Cd, Pb, Zn	<i>Zea mays</i> *	[74]
As, Cd	pigeon pea (<i>Cajanus Cajan</i>)**	[39]
Cu, Cd	Rice (<i>Oryza sativa</i> L.)**	[65]
Pb	Lantana (<i>Lantana camara</i> L.)**	[6]
	Lentil (<i>Lens culinaris</i> Medic.)**	[125]
Cd	Castor (<i>Ricinus communis</i>)*	[52]
	Alfalfa (<i>Medicago sativa</i> L.)**	[40]
Hg	<i>Populus deltoides</i> *	[22]
Se	<i>Brassica juncea</i> , <i>Astragalus bisulcatus</i> *	[15]
Zn	<i>Populus canescens</i> *	[16]

Source: Dixit *et al.* [31] *; Sumiahadi and Acar, [112] **

Plants have a range of potential mechanisms at cellular level that might be involved in the detoxification and tolerance to heavy metal stress. These are all involved in preventing toxic amounts from accumulating at sensitive sites in the cell, thereby preventing harmful effects [46]. When metals build up in tissues, they induce toxicity both directly and indirectly by destroying cell structure and replacing critical nutrients [113].

Heavy metal build-up can be avoided in a variety of ways [46, 75]. Metal accumulation can be avoided by restricting metal movement to roots with the help of mycorrhizal fungi. Huang *et al.* [52], for example, described a Zn exclusion strategy in arbuscular mycorrhizal fungi associated with *Zea mays*. According to Marques *et al.* [67], reducing the influx through the plasma membrane as well as binding to cell wall and root exudates could be viable avoidance methods for Zn retention in *Solanum nigrum* cell walls. Plants may also use other strategies to avoid metal build-up, such as stimulating

metal efflux into the apoplast. Benaroya *et al.* [12] demonstrated that this stimulation happened and that the apoplastic accumulation of Pb in *Azolla filiculoides* was significant, as well as the chelation of different ligands in the cytosol. Metal detoxification abilities in plants are aided by ligands such as phytochelatins and metalothions, as demonstrated by the engineered *Nicotiana tabacum* [75]. The transport and buildup of metals in the vacuole is one proposed avoidance tactic. *Thlaspi goesingense*, for example, improves Ni tolerance by transferring and compartmentalizing most of the internal leaf Ni into the vacuole [59].

2.2. Principles and Promises of Fundamental Phytoremediation Processes

Generally, a number of phytoremediation strategies are possible with distinct mechanism of action for the remediation of metal-polluted soils [69, 122]. Phytostabilization, in which plants are utilized to stabilize rather

than clean contaminated soil, is one of them. Phytovolatilization, in which plants extract certain metals from soil and then release them into the atmosphere [124]; phytoextraction, in which plants absorb metals from soil and translocate them to harvestable shoots, where they

accumulate [61]; phyto-filtration, which includes rhizo-filtration (use of plant roots), blasto-filtration (use of seedlings), and caulo-filtration (use of excised plant shoots) [76]. and phyto-degradation, which involves the breakdown or modification of metals within tissues by enzymes [7].

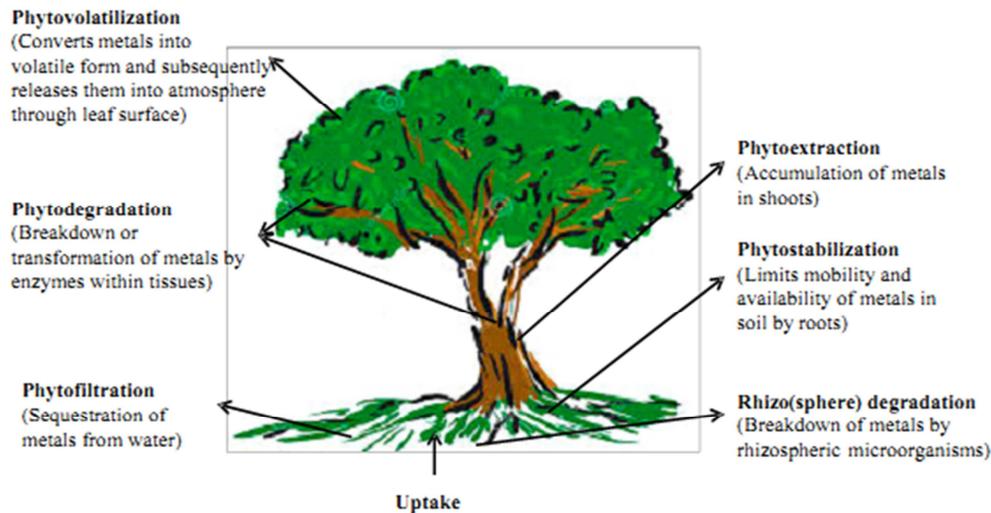


Figure 1. Various processes of heavy metals' phytoremediation [31].

2.2.1. Phyto-stabilization of Heavy Metals

Phyto-stabilization refers to grow plants on soils polluted by heavy metals [64] with principal purpose to stabilize the contaminants by limiting their mobility [5] and bioavailability [64] via the plant roots [4]. For effective stabilization, plants should be resistant to heavy metals [64]. It is predominantly applicable for remediation of heavy metals including Ar, Cr, Cd, Zn and Cu [62]. The research of Smith and Bradshaw [110], led to the development of two cultivars of *Agrostis tenuis* Sibth. and one *Festuca rubra* L. which are used for phyto-stabilization of Pb, Zn, and Cu.

Phyto-stabilization can help in a variety of circumstances involving large areas of surface contamination [26]. Phyto-stabilization is not a possibility in some highly contaminated locations since plant growth and survival are impossible [14]. Phyto-stabilization has several advantages over other soil-remediation techniques, including cost, environmental impact, ease of implementation, and aesthetic appeal [14, 101]. It also minimizes soil erosion and heavy metal migration to far-flung locations [90].

2.2.2. Phyto-extraction of Heavy Metals

Phyto-extraction, the most commonly recognized of all phytoremediation technologies, is predominantly applicable for the clean-up of polluted soils [4]. It concerned with to the use of hyper-accumulator plants [70] that are adapted to uptake large quantities of heavy metals from soil through their roots [19] and subsequently, translocate them to the above ground portions [5] where they can be stored [31] in large concentrations. As a result, the plant biomass is increased [19] and then the above-ground portions are harvested and removed so as to assure permanent deletion

of metals from the site [73]. However, in some cases, the disposal of contaminated material may become an issue. As a result, some studies propose burning of harvested plant tissue, which drastically minimizes the volume of waste that must be disposed of [61]. If important, valuable metals can be recovered from the metal-rich ash and serve as a source of revenue, thereby compensating the expense of cleanup [24, 25].

Phyto-extraction should be seen as a long-term remediation activity that will take many cropping cycles to achieve acceptable metal concentrations [61]. The amount of time required for remediation varies depending on the kind and extent of metal contamination, the length of the growing season, and the efficiency with which plants remove metal; however, it typically takes 1 to 20 years [17, 61]. Because plant growth is not sustained in extremely polluted soils, this approach is suited for the restoration of broad areas of land that are damaged at shallow depths with low to moderate amounts of metal pollutants [17, 61]. Metals in the soil should also be bioavailable, or able to be absorbed by plant roots.

Pollutants that dissolve in water are easily extracted by plants [5]. In the usual range of soil pH, for example, Pb is very insoluble and unavailable for plant uptake. As a result, plants growing in extremely contaminated locations frequently has shoots with less than 50 mg Pb g⁻¹ [26]. When grown in Pb-contaminated soil, even plants with the genetic capacity to accumulate Pb, such as *B. juncea*, will not have much Pb in their roots or shoots. The finding that certain soil-applied chelating chemicals dramatically accelerate the translocation of heavy metals, particularly Pb, from soil into shoots, provided the answer to the metal availability problem [18].

For the process to be efficient, plant should be able to tolerate high metal concentrations [70] and be efficient at translocating them from roots to the harvestable above-ground portions of the plant [17]. Furthermore, the plants are desirable to cope with difficult soil conditions (soil pH, salinity, soil structure, water content, etc) plus disease and insect problems. In fact, the success of phyto-extraction is inextricably linked to two key plant features. These abilities include the ability to rapidly create enormous amounts of biomass and the ability to accumulate high amounts of heavy metals in shoot tissues [18, 71]. Brassica juncea, while having one-third the Zn concentration in its tissue, is more successful in removing Zn from soil than *Thlaspi caerulescens*, a recognized hyper-accumulator of Zn, according to Ebbs et al. [34]. The fact that *B. juncea* produces ten times more biomass than *T. caerulescens* contributed to this advantage.

Adsorption: Root surfaces with enormous surface areas and high-affinity chemical sensors evolved specifically to absorb elemental contaminants from soils [30] and many elemental contaminants attach to root surfaces during the adsorption process [48, 97]. For example, Indian mustard (*Brassica juncea*) can rapidly concentrate Cd^{2+} , Ni^{2+} , Pb^{2+} , and Sr^{2+} into root tissues at 500 times the amounts seen in the liquid media in which it grows [97]. Sunflower roots concentrate uranium from water contaminated with low but highly dangerous levels of this oxyanion by a factor of 30,000 [33]. Similarly, tobacco roots treated to low concentrations (1–5ppm) of ionic mercury ($\text{Hg}[\text{II}]$) in liquid medium reduced the medium's $\text{Hg}[\text{II}]$ content roughly 100-fold in just a few hours [48]. Because root surfaces compete for nutrients with a variety of particulate

soil components, these adsorption mechanisms in soils are orders of magnitude less efficient than in liquid medium [89].

Uptake and Translocation: The plant cell plasma membrane contains a variety of specialized proteins involved in ion uptake and transport. These include proton pumps (ATPases that use energy and generate electrochemical gradients), co- and anti-transporters (proteins that employ the electrochemical gradients formed by ATPases to induce active ion uptake), and channels are all examples of these (proteins that facilitate the transport of ions into the cell). The interplay of ionic species during the ingestion of various heavy metal pollutants is a fundamental issue. Because root biomass cannot be harvested, translocation into shoots is desirable after uptake by roots. Metal ions are transferred from roots to shoots in a variety of ways, but little is known about them [119].

Most organic chelators, in contrast to citrate, promote metal ion absorption and translocation in plants. Plants release phytosiderophores like mugenic and avenic acids in response to metal ion shortages [35, 58]. Metal chelators boost the bioavailability of metal nutrients that are otherwise strongly bound to the soil and aid in their transport into plant tissues. Synthetic chelators can also be utilized to facilitate metal uptake and translocation. When ethylene diamine tetraacetic acid (EDTA) is applied to lead-contaminated soils, for example, the absorption and transport of lead + EDTA-chelate into stems and leaves increases 100-fold [51, 121]. In addition to citric acid's involvement in lowering aluminum uptake, plants whose secretion of specific organic acids is raised would likely show higher uptake and translocation of metal contaminants.

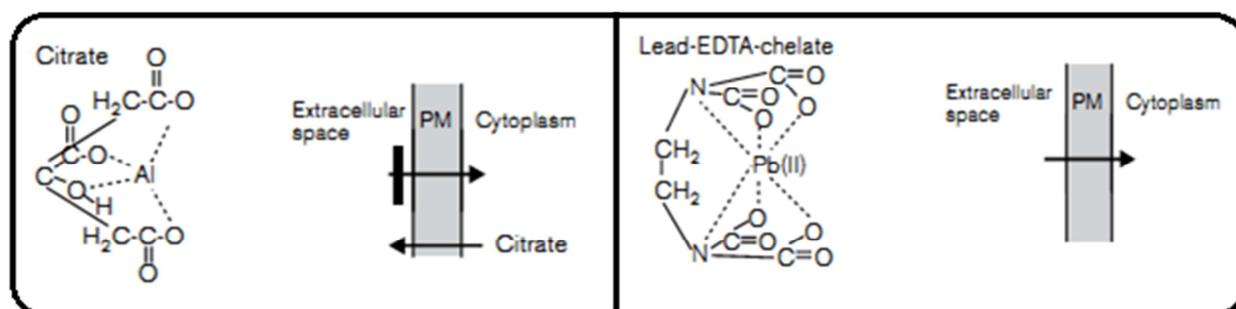


Figure 2. Metal ion uptake and translocation in plants (Richard, 2000).

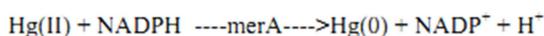
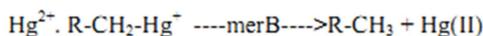
2.2.3. Phyto-volatilization of Heavy Metals

Phyto-volatilization involves the uptake of pollutants from soil into plant body [4], then transform them into low boiling [31], less harmful compounds [48], and consequently discharge them into air [31, 48] through leaves or/ and shoots [5] with the help of transpiration. Phyto-volatilization is the most contentious of all phytoremediation processes, as there is concern about the safety of releasing gaseous contaminants (As, Hg, and Se) into the atmosphere [128]. For instance, Hg is present in soil in combination with methyl group which makes it extremely toxic, but modified tobacco plants are capable to uptake and transform methyl mercury into less

toxic molecular form and ultimately release it into the atmosphere [5].

Members of the Brassicaceae are capable of emitting up to $40 \text{ g Se ha}^{-1} \text{ day}^{-1}$ as different gaseous chemicals, according to Terry et al. [116]. Cattail (*Typha latifolia* L.) is a nice example of an aquatic plant that can help in Se phytoremediation [83]. Unlike plants used for Se volatilization, those utilized for Hg volatilization are genetically engineered organisms. The bacterial organomercuriallyase (MerB) and mercuric reductase (MerA) genes have been inserted into *Arabidopsis thaliana* L. and tobacco (*Nicotiana tabacum* L.) [48, 93]. These plants take elemental mercury (II) and methyl mercury (MeHg) from the

soil and release volatile mercury (Hg(0)) into the atmosphere through their leaves [48].

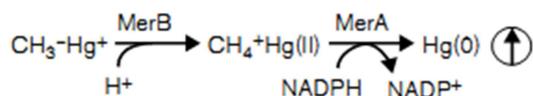


Hg (0) (elemental mercury) can be volatilized by the cell

Because the inorganic forms of these elements are eliminated and the gaseous species are unlikely to be re-deposited at or near the site, the volatilization of Se and Hg is a permanent site solution [9, 48]. Furthermore, after the first planting, places that use this technology may not require much upkeep. This type of treatment has the added advantages of causing minimal site disturbance, reducing erosion, and eliminating the need to dispose of contaminated plant debris [48, 92]. According to Heaton et al. [48], adding Hg(0) to the atmosphere would have no substantial impact on the atmospheric pool. Those who favor this strategy, however, believe that it is not appropriate to use phyto-volatilization in population centers or in areas with peculiar meteorological circumstances that encourage the rapid deposition of volatile chemicals [48, 92].

2.2.4. Phyto-degradation of Heavy Metals

In this technique, heavy metals are either broken down enzymatically or converted to less toxic forms with the aid of enzymes present in plant tissues [31]. The effectiveness of the process relies exclusively on the nature of soil, plant type and the quantity and type of pollutant to be treated [5]. After the uptake, heavy metals are subjected to various catalytical reactions within the plant living tissues. MerA and merB, two laboratory-made genes from the well-characterized bacterial mer operon, are employed in plants for mercury transformation and remediation [72]. As illustrated below, the bacterial merA gene produces a NADPH-dependent mercuric ion reductase that transforms ionic mercury (Hg(II)) to elemental mercury (Hg(0)):



Methyl mercury ($\text{CH}_3\text{-Hg}^+$) is not only the most toxic natural form of mercury, but is biomagnified efficiently in the food chain.

2.2.5. Rhizo-filtration of Heavy Metals

Plant roots are utilized to either adsorb heavy metals on roots [64], accumulate them in the root zone [5], or convert them by bacteria in the root zone [5]. Surface water, extracted ground water, and less contaminated waste water are the most common applications [4]. Land plants with deep, hairy roots are ideal for this use [21]. This approach can be used to treat large amounts of Pb and Cr [5]. Although various plants such as sunflower, Indian mustard, tobacco, rye, spinach, and corn are capable of removing lead from soil and water but sunflower has been found to remove large amount of lead after one hour of exposure [21].

3. Conclusion

Climate change, exacerbated by increased industrialization and urbanization, is causing the world to face plenty of problems and robbing it of its potential. Furthermore, toxic metal wastes produced by anthropogenic processes such as residential, municipal, agricultural, industrial, and military operations enter the soil, reducing its quality and utility. Because soil is the foundation of life, competent remediation is required. One of the current remediation options is the utilization of living organisms, notably plants (phyto-remediation). Because, in compared to other soil remediation techniques, phyto-remediation is a cost-effective and low-maintenance method that is suitable for large areas with low to moderate levels of contaminants and is environmentally friendly. Phytoremediation, on the other hand, is a long-term cleanup alternative with a variety of remediation processes. For example, air pollution may result from phyto-volatilization, whereas pollutants gathered in leaves may be released back into the ecosystem after litter fall in the case of phyto-extraction.

Furthermore, contaminants must be accessible to the plant and its root systems, despite the fact that plant life and function are influenced by a variety of edaphic and plant circumstances. As a result, future concerns should focus on modifying and improving phyto-remediation technologies that are likely to improve metal binding abilities in plant tissues and phyto-transform hazardous metals. Finally, in addition to improving and adapting diverse approaches, it is vital to prevent or avoid the discharge of dangerous substances into the environment.

References

- [1] Abhilash, M. R, Srikantaswamy S, Shiva Kumar D, Jagadish K and Shruthi L (2016). Phytoremediation of heavy metal industrial contaminated soil by *Spinacia oleracea* L. and *Zea mays* L. Int. J. Applied Sci. 4 (1): 192-99.
- [2] Abhilash, P. C.; Jamil, S. Singh, N. (2009). Transgenic plants for enhanced biodegradation and phytoremediation of organic xenobiotics. Biotechnol. Adv. 27: 474-488.
- [3] Achakzai, A. K, Bazai Z. A and Kayani S. A (2011). Accumulation of heavy metals by lettuce (*Lactuca sativa* L.) irrigated with different levels of wastewater of Quetta City Pak. J. Bot. 43 (6): 2953-60.
- [4] Akhtar, M. K, Turner N. J, Jones P. R (2013). Carboxylic acid reductase is a versatile enzyme for the conversion of fatty acids into fuels and chemical commodities. PNAS 110: 87-92.
- [5] Akshata, J. N, Udayashankara T. H, Lokesh K. S (2014). Review on bioremediation of heavy metals with microbial isolates and amendments on soil residue. International Journal of Science and Research 3: 118-123.
- [6] Alaribe, F. O and Agamuthu P (2015). Assessment of phytoremediation potentials of *Lantana camara* in Pb impacted soil with organic wasted additives Ecological Engineering 83: 513-20.

- [7] Alkorta, I.; Hernández-Allica, J. Becerril, J. M. Amezcaga, I. Albizu, I. Garbisu, C (2004). Recent findings on the phytoremediation of soils contaminated with environmentally toxic heavy metals and metalloids such as Zinc, Cadmium, Lead, and Arsenic. *Rev. Environ. Sci. Biotechnol.* 3: 71–90.
- [8] Annu, Garg A, Urmila (2016). Level of Cd in different types of soil of Rhotak district and its bioremediation. *Journal of Environmental Chemical Engineering* 4: 3797-3802.
- [9] Atkinson, R.; Aschmann, S. M. Hasegawa, D. Eagle-Thompson, E. T. and Frankenberger, J. R. W. T (1990). Kinetics of the atmospherically important reactions of dimethylselenide. *Environmental Science and Technology*, vol 24, p. 1326-1332.
- [10] Ayansina, S. A. and Olubukola, O. B (2017). A New Strategy for Heavy metal Polluted Environments: A Review of Microbial Biosorbents. *Int. J. Environ. Res. Public Health.* 14 (94): 1-16.
- [11] Belimov, A. A, Hontzas N, Safranova V. I, Demchinskaya S. V, Piluzza G, Bullitta S and Glick B R (2005). Cadmium-tolerant plant growth-promoting bacteria associated with the roots of Indian mustard (*Brassica juncea* L. Czern.) *Soil Biol. Biochem.* 37: 241-50.
- [12] Benaroya, R. O., Tzin, V., Tel-Or, E., and Zamski, E (2004). Lead accumulation in the aquatic fern *Azolla filiculoides*. *Plant Physiology and Biochemistry*, 42: 639–645.
- [13] Bermond, A., (2001). Limits of sequential extraction procedures re-examined with emphasis on the role of H⁺ reactivity. *Anal Chim Acta*, 445: 79-88.
- [14] Berti, W. R. and Cunningham, S. D (2000). Phytostabilization of metals. In: I. Raskin and B. D. Ensley eds. *Phytoremediation of toxic metals: using plants to clean-up the environment*. New York, John Wiley & Sons, Inc., p. 71-88.
- [15] Bitther, O. P.; Pilon-Smits, E. A. H.; Meagher, R. B.; Doty, S (2012). Biotechnological approaches for phytoremediation. In *Plant Biotechnology and Agriculture*; Arie Altman, A., Hasegawa, P. M., Eds.; Academic Press: Oxford, UKpp. 309–328.
- [16] Bittsanszky, A.; Kömives, T.; Gullner, G.; Gyulai, G.; Kiss, J.; Heszky, L.; Radimsky, L.; Rennenberg, H (2005). Ability of transgenic poplars with elevated glutathione content to tolerate zinc(2+) stress. *Environ. Int.* 31: 251–254.
- [17] Blaylock, M. J. and Huang, J. W (2000). Phytoextraction of metals. In: I. Raskin and B. D. Ensley eds. *Phytoremediation of toxic metals: using plants to clean-up the environment*. New York, John Wiley & Sons, Inc., p. 53-70.
- [18] Blaylock, M. J. Salt, D. E. Dushenkov, S. Zakharova, O. Gussman, C. Kapulnik, Y. Ensley, B. D. and Raskin, I (1997). Enhanced accumulation of Pb in Indian mustard by soil-applied chelating agents. *Environmental Science and Technology*, vol. 31, no. 3, p. 860-865.
- [19] Brennan, M. A, Shelley M. L. (1999). A model of the up takes translocation and accumulation of lead by maize for the purpose of phyto-extraction. *Ecological Engineering* 12: 271-297.
- [20] Bridge, G. (2004). Contested terrain: mining and the environment. *Annu. Rev. Environ. Resour.* 29: 205-259.
- [21] Camargo, F. A, Okeke, B. C, Bento, F. M, Franken-berger W. T. (2003). In-vitro reduction of hexavalent chromium by a cell-free extract of *Bacillus* sp. ES 29 stimulated by Cu²⁺. *Applied Microbiology and Biotechnology* 62: 569-573.
- [22] Che, D.; Meagher, R. B.; Heaton, A. C.; Lima, A.; Rugh, C. L.; Merkle, S. A (2003). Expression of mercuric ion reductase in Eastern cottonwood (*Populus deltoides*) confers mercuric ion reduction and resistance. *Plant Biotechnol. J.* 1: 311–319.
- [23] Chibuike, G.; Obiora, S. (2014). Heavy metal polluted soils: Effect on plants and bioremediation methods. *Appl. Environ. Soil Sci.* 1–12.
- [24] Comis, D (1996). Green remediation: Using plants to clean the soil. *Journal of soil and water conservation*, 51 (3): 184-187.
- [25] Cunningham, S. D. and Ow, D. W (1996). Promises and prospects of phytoremediation. *Plant Physiology*, 110 (3): 715-719.
- [26] Cunningham, S. D.; Berti, W. R. and Huang, J. W (1995). Phytoremediation of contaminated soils. *Trends in Biotechnology*, 13 (9): 393-397.
- [27] Dary, M., Chamber-Pérez, M. A., Palomares, A. J., Pejuelo, E. (2010). 'In situ' phytostabilisation of heavy metal polluted soils using *Lupinus luteus* inoculated with metal resistant plant-growth promoting rhizobacteria. *J. Hazard. Mater.* 177: 323-330.
- [28] Dasgupta, S, Satvat P. S and Mahinrakar A. B (2011). Ability of *Cicer arietinum* (L.) for bioremoval of lead and chromium from soil *IJTES* 2 (3): 338-41.
- [29] Dell'Amico E, Cavalva L and Andreoni V (2008). Improvement of *Brassica napus* growth under cadmium stress by cadmium-resistant rhizobacteria *Soil Biol. Biochem.* 40: 74-84.
- [30] Dittmer H. J (1995). A quantitative study of the roots and root hairs of a winter rye plant (*Secale cereale*). *Am J Bot* 24: 417-420.
- [31] Dixit, R., Wasiullah, Malaviya D, Pandiyan K, Singh UB, Sahu A, Shukla R, Singh BP, Rai JP, Sharma PK, Lade H, Paul D. (2015). Bioremediation of heavy metals from soil and aquatic environment: an overview of principles and criteria of fundamental processes. *Sustainability* 7: 2189-2212.
- [32] Dunbabin, JS, Bowmer KH (1992). Potential use of constructed wetlands for treatment of industrial waste waters containing metals. *Sci Total Environ* 111 (2.3): 151–168.
- [33] Dushenkof, S, Vasudev D, Kapulnik Y, Gleba D, Fleisher D, Ting K, Ensley B (1997). Removal of uranium from water using terrestrial plants. *Environ Sci Technol* 31: 3468-3474.
- [34] Ebbs, S. D.; Lasat, M. M.; Brandy, D. J.; Cornish, J.; Gordon, R. and Kochian, L. V (1997). Heavy metals in the environment: Phytoextraction of cadmium and zinc from a contaminated soil. *Journal of Environmental Quality*, 26: 1424-1430.
- [35] Fan, T. W, Colmer T. D, Lane A. N, Higashi R. M (1993). Determination of metabolites by 1H NMR and GC: analysis for organic osmolytes in crude tissue extracts. *Anal Biochem*, 214: 260-271.
- [36] Fashola, M.; Ngole-Jeme, V.; Babalola, O (2016). Heavy metal pollution from gold mines: Environmental effects and bacterial strategies for resistance. *Int. J. Environ. Res. Public Health*, 13, 1047.

- [37] Finnegan, P. and Chen, W (2012). Arsenic toxicity: The effects on plant metabolism. *Front. Physiol.*, 3, 182.
- [38] Fulekar, M, Singh A, Bhaduri AM. (2009). Genetic engineering strategies for enhancing phyto-remediation of heavy metals. *Afr. J. Biotechnol* 8: 529-535.
- [39] Garg, N, Singla P and Bhandari P (2014). Metal uptake, oxidative metabolism, and mycorrhization in pegeon pea and pea under arsenic and cadmium stress *Turk. J. Agric. For.* 39: 234-50.
- [40] Ghnaya, T, Mnassri M, Ghabriche R, Wali M, Poschenriender C, Lutts S and Abdelly C (2015). Nodulation by *Sinorhizobium meliloti* originated from a mining soil alleviates Cd toxicity and increases Cd-phytoextraction in *Medicago sativa* L. *Frontiers in Plant Science* 6: 1-10.
- [41] Glass, D. J (1999). U.S. and international markets for phytoremediation, 1999-2000. Needham, Mass., D. Glass Associates, p. 266.
- [42] Guar, A and A. Adholeya (2004). "Prospects of arbuscular mycorrhizal fungi in phytoremediation of heavy metal contaminated soils," *Current Science*, 86 (4): 528–534.
- [43] Guilizzoni, P (1991). The role of heavy metals and toxic materials in the physiological ecology of submersed macrophytes. *Aquat Biol* 41 (1.3): 87–109.
- [44] Gunduz, S, Uygur F. N and Kahramanoglu I (2012). Heavy metal phytoremediation potentials of *Lepidium sativum* L., *Lactuca sativa* L., *Spinacia oleracea* L. and *Raphanus sativus* L. *Herald J. Agric. Food Sci. Res.* 1 (1): 1-5.
- [45] Gurbisu, C, Alkorta I (2003). Basic concepts on heavy metal soil bioremediation. *Eur J Min Process Environ Prot* 3 (1): 58–66.
- [46] Hall, J. L. (2002). Cellular mechanisms for heavy metal detoxification and tolerance. *Journal of Experimental Botany*, 53: 1–11.
- [47] Hatano, K, Kanazawa K, Tomura K, Yamatsu T, Tsunoda K and Kubota K (2016). Molases melanoidin promotes copper uptake for radish sprouts: the potential for an accelerator of phytoextraction *Environ. Sci. Pollut. Res.* 23 (176): 56-63.
- [48] Heaton, A. C, Rugh C. L, Wang N. J, and Meagher R. B (1998). Phytoremediation of mercury and methylmercury polluted soils using genetically engineered plants. *J Soil Contam*, 7: 497-509.
- [49] Hegedusova, A, Jakabova S, Vargova A, Hegeus O and Pernyeszi T. J (2009). Use of phytoremediation techniques for elimination of lead from polluted soils *Nova Biotechnologica*, 9 (2): 125-132.
- [50] Hinchman, R. R., M. C. Negri, and E. G. Gatli (1995). "Phytoremediation: using green plants to clean up contaminated soil, groundwater, and wastewater," Argonne National Laboratory Hinchman, Applied Natural Sciences, Inc.,
- [51] Huang, J. W, Chen J, Berti W. R, Cunningham S. D (1997). Phytoremediation of lead-contaminated soils: role of synthetic chelates in lead phytoextraction. *Environ Sci Tech*, 31: 800-805.
- [52] Huang, H.; Yu, N.; Wang, L.; Gupta, D. K.; He, Z.; Wang, K.; Zhu, Z.; Yan, X.; Li, T.; Yang, X. E (2011). The phytoremediation potential of bioenergy crop *Ricinus communis* for DDTs and cadmium co-contaminated soil. *Bioresour. Technol.*, 102: 11034–11038.
- [53] Huang, Y., Chen, Y., and Tao, S. (2002). Uptake and distribution of Cu, Zn, Pb and Cd in maize related to metals speciation changes in rhizosphere. *Chinese Journal of Applied Ecology*, 13: 859-862.
- [54] Jahanbakhshi, S, Rezaei M. R and Sayyari-Zahan M. H (2014). Optimization of phytoremediation in Cd-contaminated soil by using Taguchi method in *Spinacia oleracea* Proceedings of the International Academy of Ecology and Environmental Sciences vol 4 ed W Zhang (Hongkong: International Academy of Ecology and Environmental Sciences) pp 185-93.
- [55] Jamil, S.; Abhilash, P. C.; Singh, N.; Sharma, P. N. (2009). *Jatropha curcas*: A potential crop for phytoremediation of coal fly ash. *J. Hazard. Mater.*, 172: 269–275.
- [56] Jayanthi, B, Emenike C. U, Agamuthu P, Simarani K, Mohamad S, Fauziah S. H (2016). Selected microbial diversity of contaminated landfill soil of Peninsular Malaysia and the behaviour towards heavy metal exposure. *J. of Catena* 147: 25-31.
- [57] Kambhampati, M. S and Vu V. T (2013). EDTA enhanced phytoremediation of copper contaminated soils using chickpea (*Cicer arietinum* L.) *Bull. Environ. Contam. Toxicol.* 91: 310-13.
- [58] Kinnerseely, A. M (1993). The role of phytochelates in plant growth and productivity. *Plant Growth Regul*, 12: 207-217.
- [59] Kramer, U., Pickering, I. J., Prince, R. C., Raskin, I., and Salt, D. E. (2000). Subcellular localization and speciation of nickel in hyperaccumulator and non-accumulator *Thlaspi* species. *Plant Physiology*, 122; 1343–1353.
- [60] Kulshreshtha, A, Agrawal R, Barar M, Saxena S. (2014). A Review on Bioremediation of Heavy Metals in Contaminated Water. *IOSR Journal of Environmental Science, Toxicology and Food Technology* 8: 44-50.
- [61] Kumar, P. B., Dushenkov, V., Motto, H., and Raskin, L. (1995). Phytoextraction: The use of plants to remove heavy metals from soils. *Environmental Science and Technology*, 29: 263–290.
- [62] Kunito, T, Saeki K, Oyaizu K, Mutsumoto S (2010). Characterization of copper resistant bacterial communities in copper contaminated soils. *European Journal of Soil Biology* 37: 95-102.
- [63] Lasat, M. M. Pence, N. S. Garvin, D. F. Ebbs, S. D. and Kochian, L. V (2000). Molecular physiology of zinc transport in the Zn hyperaccumulator *Thlaspi caerulescens*. *Journal of Experimental Botany*, 51 (342): 71-79.
- [64] Latha, M, Indirani R, Kamaraj S. (2004). Bioremediation of polluted soil. *Agri. Rev* 25: 252-266.
- [65] Li, P, Wang X, Zhang T, Zhou D and He Y (2008). Effect of several amendments on rice growth and uptake of copper and cadmium from a contaminated soil *J. Environ. Sci.* 20: 449-55.
- [66] Malecka, A, Piechalak A and Morkunas I (2008). Accumulation of lead in root cells of *Pisum sativum* *Acta Physiol Plant* 306: 29-37.

- [67] Marques, A. P., Oliveira, R. S., Samardjieva, K. A., Pissarra, J., Rangel, A. O., and Castro P. M. L (2007). *Solanum nigrum* in contaminated soil: Effect of arbuscular mycorrhizal fungi on zinc accumulation and histolocalisation. *Environmental Pollution*, 145: 691–699.
- [68] Marrugo-Negrete, J, Durango-Hernandez J, Pinedo-Hernandez J, Olivero-Verbel J and Diez S (2015) Phytoremediation of Hg-contaminated soils by *Jatropha curcas* *Chemosphere* 127: 58-63.
- [69] Martinez, M. Bernal, P. Almela, C. Vélez, D. García-Agustín, P. Serrano, R. Navarro-Aviñó, J (2006). An engineered plant that accumulates higher levels of heavy metals than *Thlaspi caerulescens*, with yields of 100 times more biomass in mine soils. *Chemosphere*, 64: 478-485.
- [70] Matheickal, J. T, Yu Q. (1999). Biosorption of lead (II) and copper (II) from aqueous solution by pre-treated biomass of Australian marine algae. *Biores. Technol* 69: 223-229.
- [71] McGrath, S. P (1998). Phytoextraction for soil remediation. In: Brooks, R. R., ed. *Plants that hyperaccumulate heavy metals: their role in phytoremediation, microbiology, archaeology, mineral exploration and phytomining*. New York, CAB International, p. 261-288.
- [72] Meagher, R. B, Rugh C. L, Kandasamy M. K, Gragson G, Wang N. J (2000). Engineered phytoremediation of mercury pollution in soil and water using bacterial genes. In *Phytoremediation of Contaminated Soil and Water*. Edited by Terry W, Bañuelos G. Berkeley, California: Ann Arbor Press, Inc.; 201-219.
- [73] Meenambigai, P, Vijayaraghavan R, Gowri RS, Rajarajeswari P, and Prabhavathi P. (2016). Biodegradation of Heavy Metals - A Review. *Int. J. Curr. Microbiol. App. Sci* 5: 375-383.
- [74] Meers, E.; van Slycken, S.; Adriaensen, K.; Ruttens, A.; Vangronsveld, J.; Du Laing, G.; Witters, N.; Thewys, T.; Tack, F. M (2010). The use of bio-energy crops (*Zea mays*) for “phytoattenuation” of heavy metals on moderately contaminated soils: A field experiment. *Chemosphere*, 78: 35–41.
- [75] Mejáre, M. and Bülow, L (2001). Metal binding proteins and peptides in bioremediation and phytoremediation of heavy metals. *Trends in Biotechnology*, 19 (2): 67-75.
- [76] Mesjasz-Przybyłowicz, J.; Nakonieczny, M.; Migula, P.; Augustyniak, M.; Tarnawska, M.; Reimold, W. U.; Koeberl, C.; Przybyłowicz, W.; Glowacka, E. Uptake of cadmium, lead, nickel and zinc from soil and water solutions by the nickel hyperaccumulator *Berkheya coddii*. *Acta Biol. Cracov. Bot.* 2004, 46, 75–85.
- [77] Mojiri A 2011. The potencial of corn (*Zea mays*) for phytoremediation of soil contaminated with cadmium and lead *J. Biol. Environ. Sci.* 5 (13): 17-22.
- [78] Morera, M. T., J. C. Echeverria, C. Mazkarian and J. J. Garrido, (2001). Isotherms and sequential extraction procedures for evaluating sorption and evaluation of heavy metals in soils. *Envir pollution*, 113: 135-144.
- [79] Mukhtar, B, Malik MF, Shah SH, Azzam A, Slahuddin, Liaqat I. (2017). Heavy Metal Bioremediation in Soil: Key Species and Strategies involved in the Process. *International Journal of Applied Biology and Forensics* 1 (2): 5-15.
- [80] Patel, M and Subramanian R. B (2006). Effect of a chelating agent on lead uptake by *Spinacia olearea* *Poll. Res.* 25 (1): 77-79.
- [81] Pathak, C, Chopra A. K and Zivastava S (2013). Accumulation of heavy metals in *Spinacia oleracea* irrigated with paper mill effluent and sewage *Environ. Monit. Assess.* 185 (73): 43-52.
- [82] Pilon-Smits, E. (2005). Phytoremediation. *Annual Revisions in Plant Biology*, 56: 15–39.
- [83] Pilon-Smits, E. A. H.; Desouza, M. P.; Hong, G.; Amini, A.; Bravo, R. C.; Payabyab, S. T. and Terry, N (1999). Selenium volatilization and accumulation by twenty aquatic plant species. *Journal of Environmental Quality*, 28 (3): 1011-1017.
- [84] Prabhu, S.; Poulouse, E. K (2012). Silver nanoparticles: Mechanism of antimicrobial action, synthesis, medical applications, and toxicity effects. *Int. Nano Lett.*, 2: 1–10.
- [85] Qian, H.; Peng, X.; Han, X.; Ren, J.; Sun, L.; Fu, Z (2013). Comparison of the toxicity of silver nanoparticles and silver ions on the growth of terrestrial plant model *Arabidopsis thaliana*. *J. Environ. Sci.*, 25: 1947–1956.
- [86] Quainoo, A. K, Konadu A and Kumi M (2015). The potential of shea nut shells in phytoremediation of heavy metals in contaminated soil using lettuce (*Lactuca sativa*) as a test crop *J. Bioremed. Biodeg.* 6 (1): 1-7.
- [87] Rakhshae, R M. Giahi, and A. Pourahmad (2009). “Studying effect of cell wall’s carboxyl-carboxylate ratio change of *Lemna minor* to remove heavy metals from aqueous solution,” *Journal of Hazardous Materials*, 163 (1): 165–173.
- [88] Rashid, A, Mahmood T, Mehmood F, Khalid A, Saba B, Batoool A and Riaz A (2014). Phytoaccumulation, competitive adsorption and evaluation of chelators-metal interaction in lettuce plant *Environ. Eng. Management J.* 13 (10): 2683-92.
- [89] Raskin I, Smith R. D, Salt D. E (1997). Phytoremediation of metals: using plants to remove pollutants from the environment. *Curr Opin Biotechnol*, 8: 221-226.
- [90] Raskin, I. and Ensley, B. D (2000). *Phytoremediation of toxic metals: using plants to clean up the environment*. New York, John Wiley and Sons, 352 p. ISBN 0-47-119254-6.
- [91] Richard, B. M (2000). *Phytoremediation of toxic elemental and organic pollutants*. Elsevier Science Ltd. 3: 153–162.
- [92] Rugh, C. L.; Bizily, S. P. and Meagher, R. B (2000). Phytoreduction of environmental mercury pollution. In: Raskin, I. and Ensley, B. D., eds. *Phytoremediation of toxic metals: using plants to clean- up the environment*. New York, John Wiley and Sons, p. 151-170.
- [93] Rugh, C. L.; Gragson, G. M.; Meagher, R. B. and Merkle, S. A (1998). Toxic mercury reduction and remediation using transgenic plants with a modified bacterial gene. *Hortscience*, 33 (4): 618-621.
- [94] Ruttens, A.; Boulet, J.; Weyens, N.; Smeets, K.; Adriaensen, K.; Meers, E.; van Slycken, S.; Tack, F.; Meiresonne, L.; Thewys, T.; et al (2011). Short rotation coppice culture of willows and poplars as energy crops on metal contaminated agricultural soils. *Int. J. Phytorem.*, 13: 194–207.
- [95] Salaskar, D, Shrivastava M and Kale S. P (2011). Bioremediation potential of spinach (*Spinacia oleracea* L.) for decontamination of cadmium in soil *Current Sci.* 101 (10): 1359-1363.

- [96] Salem, H. M, Eweida E. A, Farag A. (2000). Heavy metals in drinking water and their environmental impact on human health. In ICEHM 2000: Cairo University: Giza, Egypt. 542-556.
- [97] Salt, D. E, and Kramer U (1999). Mechanisms of metal hyperaccumulation in plants. In *Phytoremediation of Toxic Metals: Using Plants to Clean-up the Environment*. Edited by Raskin I, Ensley BD. New York: John Wiley and Sons; 231-246.
- [98] Salt, D. E.; Smith, R. D. and Raskin, I (1998). Phytoremediation. *Annual Review of Plant Physiology and Plant Molecular Biology*, 49: 643-668.
- [99] Samarghandi, M. R, Nouri J, Mesdaghinia A. R, Mahvi A. H, Nasser S, Vaezi F (2007). Efficiency removal of phenol, lead and cadmium by means of UV/TiO₂/H₂O₂ processes. *Int J Environ Sci Technol* 4 (1): 19–25.
- [100] Schmoeger, M. E., Owen, M., and Grill, E. (2000). Detoxification of arsenic by phytochelatin in plants. *Plant Physiology*, 122: 793–801.
- [101] Schnoor, J. L (2000). Phytostabilization of metals using hybrid poplar trees. In: RASKIN, I. and ENSLEY, B. D., eds. *Phytoremediation of toxic metals: using plants to clean-up the environment*. New York, John Wiley & Sons, Inc., p. 133-150.
- [102] Schnoor, J. L. Light, L. A. McCutcheon, S. C. Wolfe, N. L. and Carreira, L. H (1995). Phytoremediation of organic and nutrient contaminants. *Environmental Science and Technology*, 29 (7): 318-323.
- [103] Sharma, H (2016). Phytoremediation of lead using *Brassica juncea* and *Vetiveria zizanioides* *Int. J. Life Sci. Res.* 4 (1): 91-96.
- [104] Sharma, S, Sharma P and Mehrotra P (2010). Bioaccumulation of heavy metals in *Pisum sativum* L. growing in fly ash amended soil *J. American Sci.* 6 (6): 43-50.
- [105] Sharma, R. K., Agrawal, M. (2006). Single and combined effects of cadmium and zinc on carrots: uptake and bioaccumulation. *J. Plant Nutri.* 29, 1791-1804.
- [106] Shazia, I, Uzma, Sadia GR, Talat A. (2013). Bioremediation of heavy metals using isolates of filamentous fungus *Aspergillus fumigatus* collected from polluted soil of Kasur, Pakistan. *International Research Journal of Biological Sciences* 2: 66-73.
- [107] Sheng, X. F and Xia J. J (2006). Improvement of rape (*Brassica napus*) plant growth and cadmium uptake by cadmium-resistant bacteria *Chemosphere* 64: 1036-42.
- [108] Shtangeeva, I, J. V. Laiho, H. Kahelin, and G. R. Gobran (2004). "Phytoremediation of metal-contaminated soils. Symposia Papers Presented Before the Division of Environmental Chemistry," American Chemical Society, Anaheim, Calif, USA,
- [109] Singh, A and Fulekar M. H (2012). Phytoremediation of heavy metals by *Brassica juncea* in aquatic and terrestrial environment. *The Plant Family Brassicaceae: Contribution Towards Phytoremediation* ed N. A Anjum, I. Ahmad, M. E Pereira, A. C Duarte, S Umar and N. A Khan (Amsterdam: Springer Science+Business Media) pp 153-69.
- [110] Smith, R. A. and Bradshaw, A. D (1992). Stabilization of toxic mine wastes by the use of tolerant plant populations. *Transactions of the Institution of Mining and Metallurgy*, 81: 230-237.
- [111] Smolinska, B and Szczodrowska A (2016). Antioxidative response of *Lepidium sativum* L. during assisted phytoremediation of Hg contaminated soil *New Biotechnology*.
- [112] Sumiahadi, A and R Acar (2018). A review of phytoremediation technology: heavy metals uptake by plants. *IOP Conf. Ser.: Earth Environ. Sci.* 142: 1755-1315.
- [113] Taiz, L., and Zeiger, E. (2002). *Plant physiology*. 3rd ed. Sunderland, Mass.: Sinauer Associates Inc.
- [114] Takeda, R, Sato Y, Yoshimura R, Komemushi S and Sawabe A. (2006). Accumulation of heavy metals by cucumber and *Brassica juncea* under different cultivation conditions *Proc. Ann. Int. Conf. on Soil Sediments Water Energy (Massachusetts) 11 (California: The Berkeley Electronic Press)* pp 293-99.
- [115] Tchounwou, P. B, Yedjou C. G, Patlolla A. K, Sutton D. J. (2014). Heavy metal toxicity and the environment. *PMC* 101: 133-164.
- [116] Terry, N.; Carlson, C.; Raab, T. K. and Zayed, A (1992). Rates of selenium volatilization among crop species. *Journal of Environmental Quality*, 21: 341-344.
- [117] Tiecher, T, Ceretta C. A, Ferreira P. A. A, Lourenzi C. A, Tiecher T, Girotto E, Nicoloso F. T, Soriani H. H, De Conti L, Mimmo T, Cesco S and Brunetto G (2016). The potential of *Zea mays* L. in remediating copper and zinc contaminated soils for grapevine production *Geoderma* 262: 52-61.
- [118] Turan, M and Estringu A (2007). Phytoremediation based on canola (*Brassica napus* L.) and Indian mustard (*Brassica juncea* L.) planted on spiked soil by aliquot amount of Cd, Cu, Pb, and Zn *Plant Soil Environ.* 53 (1): 7-15.
- [119] USDE (U.S. Department of Energy) (1994). "Plume Focus Area, December. Mechanisms of plant uptake, translocation, and storage of toxic elements. Summary Report of a workshop on phytoremediation research needs,"
- [120] Vangronsveld, J. and Cunningham, S. D (1998). *Metal-contaminated soils: in-situ in activation and phytoremediation*. Springer-Verlag, Berlin, Heidelberg, 265 p.
- [121] Vassil, A. D, Kapulnik Y, Raskin I, Salt D. E (1998). The role of EDTA in lead transport and accumulation by Indian mustard. *Plant Physiol*, 117: 447-453.
- [122] Vidali, M., (2001). Bioremediation. An overview. *Pure appl. Chem.* 73: 1163-1172.
- [123] Volk, T. A.; Abrahamson, L. P.; Nowak, C. A.; Smart, L. B.; Tharakan, P. J. (2006). White, E. H. The development of short-rotation willow in the northeastern United States for bioenergy and bioproducts, agroforestry and phytoremediation. *Biomass Bioener.*, 30: 715–727.
- [124] Vroblesky, D. A., Nietch, C. T., and Morris, J. T. (1999). Chlorinated ethanes from ground water in tree trunks. *Environmental Science and Technology*, 33: 510–515.
- [125] Wani, P. A and Khan M. S 2012 Bioremediation of lead by a plant growth promoting Rhizobium species RL9 *Bacteriology J.* 2 (4): 66-78.

- [126] Wani, P. A, Khan M. S and Zaidi A (2007). Impact of heavy metal toxicity on plant growth, symbiosis, seed yield and nitrogen and metal uptake in chickpea Australian J. Exp. Agric. 47: 712-20.
- [127] Wani, P. A, Khan M. S and Zaidi A (2008). Effects of heavy metal toxicity on growth, symbiosis, seed yield and metal uptake in pea grown in metal amended soil. Bull. Environ. Contam. Toxicol. 81: 152-58.
- [128] Watanabe, M. E (1997). Phytoremediation on the brink of commercialization. Environmental Science and Technology, 31: 182-186.
- [129] Wright, D. J, Otte M. L (1999). Plant effects on the biogeochemistry of metals beyond the rhizosphere. Biol Environ Proc R Ir Acad 99B (1): 3-10.
- [130] Wuana, R. A, Okieimen FE. (2011). Heavy metals in contaminated soils: A review of sources, chemistry, risks and best available strategies for remediation. ISRN Ecol. Article 20.
- [131] Yadav, S. K, Juwarkar A. S, Kumar P, Thawale P. R, Singh S. K and Chakrabarti T (2009). Bioaccumulation and phyto-translocation of arsenic, chromium, and zinc by *Jatropha curcas* L.: impact of dairy sludge and biofertilizer Biosource Tech. 100 (46): 16-22.